ATTEMPTED MICROBIOLOGICAL PRODUCTION OF ω-FLUOROAVENACIOLIDE BY ASPERGILLUS AVENACEUS

BRIAN E. CROSS[®] and PAUL HENDLEY

Department of Organic Chemistry, The University, Leeds LS2 9JT, England.

(Received in UK 20 October 1977; Accepted for publication 31 October 1977)

Abstract—12-Fluoro-3-oxododecanoic acid has been synthesised; neither this acid nor 10-fluorodecanoic acid were converted into fluoroavenaciolide by cultures of Aspergillus avenaceus.

FOLLOWING our successful microbiological production of two fluorogibberellins,¹ we have attempted to utilise similar principles in the preparation of fluorinated analogues of polyketide derived mould products (see Ref. 2). Aspergillus avenaceus G. Smith produces the antifungal metabolite³ avenaciolide (1) which is reported to be biosynthesised from 3-oxododecanoic and succinic acids,⁴ and appeared therefore to be an appropriate organism for our experiments.

It seemed reasonable to assume that the end of the long alkyl chain of 3-oxododecanoic acid plays a relatively minor role in the biosynthesis of avenaciolide. On this basis, 12-fluoro-3-oxododecanoic acid was chosen as a suitable analogue of this precursor for feeding to cultures of *A. avenaceus*. The 12-fluoro-keto-acid was prepared from 10-fluorodecanoic acid³ by the sequence shown in Fig. 1 (see Refs 6-8) and was found to be stable in the culture medium. An attempt to hydrolyse the ester 3 in acetic acid solution⁷ gave 11-fluoro-2-oxoundecane.



Still cultures of A. avenaceus were fed with 12-fluoro-3-oxododecanoic acid (Table 1) in ethanol and dimethylsulphoxide solutions. The yields of avenaciolide in both the fed and solvent-blank cultures were greatly reduced and abnormal white growths appeared on the top of the mycelial mats. Subsequently investigations revealed that in contrast to many other fungi,⁹ A. avenaceus would only grow normally if the concentration of ethanol was below 1.5%, but the fungus was unaffected by relatively large concentrations of sodium fluoroacetate (Table 1). The small amounts of avenaciolide from these feeds



Fig. 1.

Tab	le 1.	Feeding	experiment	s with A	арепасенз
-----	--------------	---------	------------	----------	-----------

Substrate		Solvent		No. of flashs	Tield of orade avanaiolide	Yield of pure avenaciolide	Ricrossiyses		
Cospound	mg/flask		nl/flask	fed	ng/flask	ng/flask	Found:		_
12-Fluoro-3- oxododecancic acid	25	BFOR	2•5	8		trace	 .	•	
12-Fluoro-3-cro- dodecencic acid	30	Ne ₂ 80	4	5	~12	3	_		
Ethyl 12-fluoro- 5-axododeanaate	50	BP0E	0-5	3	60	—	0, 67- 85; I, 8-3;	7, 0 6	•
10-Fluorodecanois acid	50	EVOE	4	1	16	9	C, 67-5; I, 8-55;	7,06	
10-Fluorodecanoic acid	50	BLOE	0-5	3	38	_	C, 67-85: I, 8-5;	7, OK	
Sodium fluoro- acetate	3-80	H20	0-25-6	6	20	13	0, 67• 85; I, 8-4;	7, 0 6	

* Calo. for C15H22O4: C, 67-6; H, 8-34

were examined by NMR and mass spectroscopy, and where possible by microanalysis, but no evidence for the presence of the ω -fluoro-analogue (2) was obtained. Ethyl 12-fluoro-3-oxododecanoate (3) was also fed (Table 1) since it was possible that the ester might enter the cells and then undergo hydrolysis to give the keto-acid *in* situ; a good yield of avenaciolide was obtained, but it contained no detectable amount of the fluoro-analogue.

If 3-oxododecanoic acid is a precursor of avenaciolide then it is reasonable to assume that decanoic acid might act as a "starter" unit (see Ref. 10). Consequently 10fluorodecanoic acid was fed to the fungus (Table 1) and the avenaciolide was isolated, but again there was no evidence that ω -fluoroavenaciolide had been produced. It was concluded that A. avenaceus is probably incapable of utilising the fluoro-acids for the production of significant amounts of fluoroavenaciolide (2).

EXPERIMENTAL

Details of chromatographic materials and conditions used for the determination of physical data, etc. have been reported.¹

10-Fluorodecanoic acid. Prepared by the literature method,⁵ the acid, m.p. 49-49.5°, showed τ 7.60 (2H, t, J 6.5 Hz, 2-H₂) and 5.51 (2H, dt, J 47 and 6 Hz, 10-H₂); ϕ^{\bullet} 218.7 (7 lines, 10-F).

Ethyl 2-acetyl-12-fluoro-3-oxododecanoate and ethyl 12fluoro-3-oxododecanoate. 10-Fluorodecanoyl chloride, prepared by treating the acid (5.0 g) with thionyl chloride in benzene under reflux followed by evaporation in vacuo was a gum, v_{max} (film) 1800 cm⁻¹.

Ethyl acetoacetate (3.9 g) in benzene (30 m) was treated with Na (605 mg) and then the above acid chloride in-benzene was added dropwise. The mixture was refluxed for 15 min, poured onto ice, and acidified with H₂SO₄ (5%), EtOH (25 mi) was added, the layers were separated, and the aqueous phase was extracted with benzene. The combined extracts were washed with H₂O-EtOH (9:1); recovery afforded an orange oil (8.1 g) containing ethyl 2-acetyl-12- fluoro-3-oxododecanoate.

The oil and Na (650 mg) in EtOH (20 ml) were left at room temp. for 12 hr and were poured onto ice and acidified with 2N H₂SO₄. Recovery in ether gave a red gum (7.7 g) which was chromatographed on silica gel (24 × 4 cm). Elution with EtOAc-light petroleum (1:49) gave unchanged ethyl 2-acetyl-12-fluoro-3oxododecanoate as an oil (Found: m/e 302.1896. C₁₆H₂₇FO₄ requires; M, 302.1893), ν_{max} 1740 br; τ 7.63 (3H, s, COMe), 7.65 (2H, t, J 6.5 Hz, 4-H₂), 5.82 (2H, q, J 6.5 Hz, OCH₂Me) and 5.55 (2H, dt, J 47 and 6.5 Hz, 12-H₂).

Elution with EtOAc—light petroleum (1:19) afforded ethyl-12fluoro-3-oxododecanoate (3; 1.71 g) which crystallised at low temp. as rosettes, m.p. 27-29° (Found: m/e 260.1789. $C_{14}H_{25}FO_3$ requires: M, 260.1788), ν_{max} 3420, 1742, 1720, 1650 and 800 cm⁻¹; τ 7.45 (2H, t, J 6 Hz, 4-H₂), 6.57 (2H, 's, 2-H₂), 5.77 (2H, q, J 7 Hz, OCH₂Me) and 5.52 (2H, dt, J 47 and 6 Hz, 12-H₂).

Further elution gave 10-fluorodecanoic acid.

12-Fluoro-3-oxododecanoic acid. The crude ester (200 mg) from the preceding experiment was stirred with NaOH soln (1%; 30 ml) at room temp. for 20.5 hr. Careful acidification with dil. HCl precipitated 12-fluoro-3-oxododecanoic acid which was

 \dagger Avenaciolide gives a small molecular ion, but the M + 1 ion is much more intense.

collected by filtration, m.p. (rapid heating) 73-75° (decomp.) (Found: C, 61.3; H, 9.05; F, 8.95%; m/e 232. $C_{12}H_{21}FO_3$ requires: C, 62.0; H, 9.1; F, 8.2%; M, 232), ν_{max} 3200 br, 1725 and 1700 cm⁻¹; τ 7.31 (t, J 7 Hz, 4-H₂), 6.46 (s, 2-H₂) and 5.55 (2H, dt J 48 and 6 Hz, 12-H₂). (All samples of the keto-acid contained small amounts of 11-fluoroundecan-2-one).

The keto-acid was recovered after an ethanolic soln (23 mg in 2.5 ml) was added to sterile culture medium (200 ml) and left to stand for 50 hr.

Its methyl ester, prepared with diazomethane, was a solid (Found: m/e 246.1631. C₁₃H₂₂FO₃ requires: M, 246.1624).

11-Fluoroundecan-2-one. Ethyl 12-fluoro-3-oxododecanoate (1.55 g) in glacial HOAc (30 ml) was stirred at room temp. and conc. HCl was added until the soln became turbid. After 60 hr the soln was evaporated *in vacuo* and the product was recovered in ether. Chromatography on silica gel (20 × 2 cm) and elution with EtOAc---light petroleum (1:39) yielded 11-fluoroundecan-2-one (955 mg) as an oil (Found: m/e 188.1578. C₁₁H₂₁FO requires: M, 188.1576), ν_{max} (film) 1720 cm⁻¹; τ 7.9 (3H, s, 1-H₃), 7.61 (2H, t, J 7 Hz, 3-H₂) and 5.56 (2H, dt, J 48 and 6 Hz, 11-H₂).

Feeding experiments. Aspergillus avenaceus G. Smith (C.M.I. 16140) was grown as a surface culture³ on Czapek-Dox medium (200 ml) in 1 lpenicillin vessels at 26°. The fluoro-acids were dissolved in the appropriate solvent, sterilised by means of a Seitz filter, and added to 6-7 day old cultures. The flasks were harvested after a further 7-8 days and the avenaciolide was isolated by the literature method.³ The yield of crystalline avenaciolide from control flasks increased, during the course of the investigation, from ca. 20 to ca. 80 mg/flask. Blank cultures were treated with sterile solvent only.

Avenaciolide from fed cultures (see Table 1) was crystallised once and then examined by TLC, IR, NMR, and where possible by microanalysis for fluorine. All the samples were subjected to mass spectroscopy with the multiplier set at greatly increased gain in a search for ions at m/e 284 and 2851 ($C_{15}H_{21}FO_4$ requires M, 284). In every case the avenaciolide from fed cultures was indistinguishable from a pure reference sample.

Acknowledgements—We thank Mr. T. R. Etherington for technical assistance and the S.R.C. for a Research Studentship (to P.H.).

REFERENCES

- ¹J. H. Bateson and B. E. Cross, J. Chem. Soc. Perkin I, 1131 (1974).
- ²B. E. Cross and P. Hendley, *Ibid.* Perkin I, 2523 (1975).
- ³D. Brookes, B. K. Tidd and W. B. Turner, *Ibid.* 5385 (1963).
- ⁴M. Tanabe, T. Hamasaki, Y. Suzuki and L. F. Johnson, *Ibid.* Chem. Comm. 212 (1973).
- ⁵F. L. M. Pattison, J. B. Stothers and R. G. Woolford, J. Am. Chem. Soc. 78, 2255 (1956).
- S. Ställberg-Stenhagen and E. Stenhagen, Arhiv Kemi, Mineral, Geol. A20, No. 19 (1945).
- ⁷M. A. Mitz, A. E. Axelrod and K. Hofmann, J. Am. Chem. Soc. 72, 1231 (1950).
- Y. Asahine and S. Nakayama, J. Pharm. Soc. Japan, 526, 1058 (1925).
- ⁹J. W. Browne, W. A. Denny, E. R. H. Jones, G. D. Meakins, Y. Morisawa, A. Pendlebury and J. Pragnell, J. Chem. Soc. Perkin
- I, 1493 (1973) and earlier papers.
- ¹⁰E. Leete and J. O. Olson, J. Am. Chem. Soc. 94, 5472 (1972).